

Sandhoff Disease is Corrected in a Mouse Model by scAAV9-HEXM Gene Transfer

BM Quinville¹, AE Ryckman¹, NM Deschenes¹, M Mitchell², K Singh², JG Keimel³, WF Kaemmerer³, and JS Walia^{1,2,4}

¹Centre for Neuroscience Studies, Queen's University, Kingston, Ontario, K7L 3N6, Canada; ²Department of Pediatrics, Queen's University, Kingston, Ontario, K7L 2V7, Canada; ³New Hope Research Foundation, North Oaks, Minnesota, USA;

Infantile

 \sim 4 years.

Juvenile

adulthood.

adulthood.

Adult

Onset: ~ 6 months of age; death:

Onset: 3-10 years, may survive to

<2% residual HexA activity.

Most prevalent form of SD

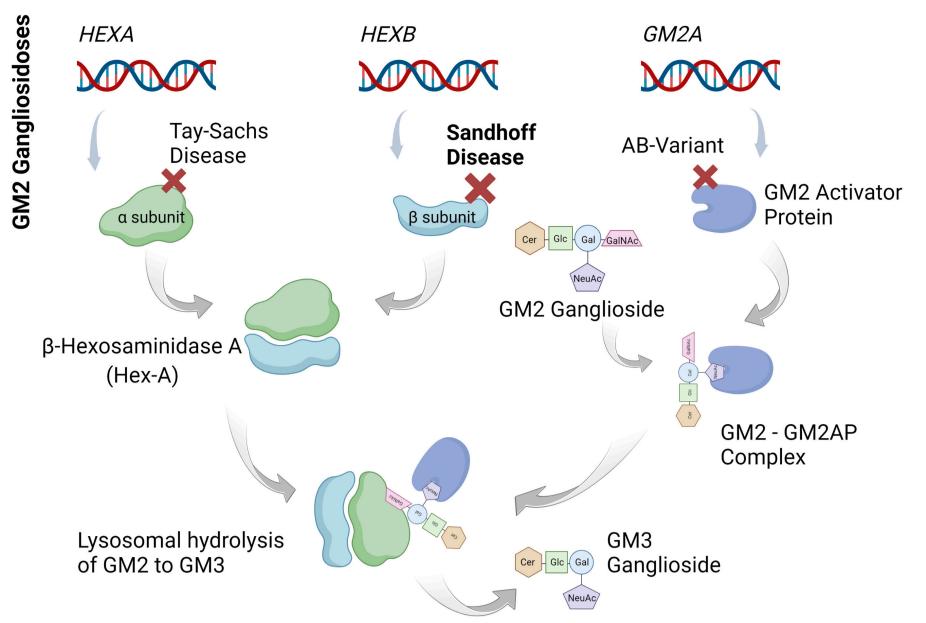
2-10% residual HexA activity.

Onset: Adolescence/ early

Slowest progression of the three.

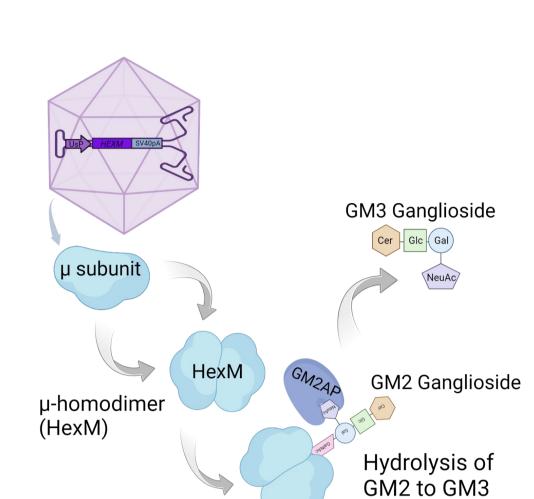
7-10% residual HexA activity.

⁴Department of Biomedical and Molecular Sciences, Queen's University, Kingston, Ontario, K7L 3N6, Canada



Sandhoff disease (SD) is caused by the excessive accumulation of GM2 gangliosides in the lysosomes of neuronal cells. Typically, these lipids are hydrolyzed by β -hexosaminidase A (Hex-A), a heterodimer comprised of an α and a β-subunit. Mutations in the gene encoding either subunit can lead to improper functioning of the enzyme. SD is caused by a mutation in the HEXB gene resulting in a deficient or absent β-subunit and subsequent accumulation of GM2 gangliosides. This causes widespread cell death, and consequently progressive symptoms and rapid neurological decline culminating in death.

A homodimer formed by a novel hybrid μ-subunit called HexM, an isoenzyme of human Hex-A, has been recently developed and shown to hydrolyze GM2 gangliosides in vivo¹. Previous studies have determined the effectiveness of gene transfer with the gene, HEXM, packaged in a self-complementary adenoassociated viral vector, serotype 9 (scAAV9), through increased life span in a SD mouse model $(Hexb^{(-/-)})^2$.



This study aims to determine the dose response of the scAAV9-HEXM treatment in the SD mouse model through dual delivery of treatment via intra-cisterna magna (ICM) and intravenous (IV) routes, along with the ancillary administration of immunosuppressant drugs.

Cohort & Genotype	N	Treatment Group	Dose via ICM infusion	Dose via IV infusion	Total dose
Heterozygous	8	Vehicle	0	0	0
Hexb ^(-/-)	9	Vehicle	0	0	0
Hexb ^(-/-)	9	ICM low	1.0e11 vg	0	1.0e11 vg
Hexb ^(-/-)	10	ICM high	2.5e11 vg	0	2.5e11 vg
Hexb ^(-/-)	9	Dual: ICM high + IV low	2.5e11 vg	2.5e11 vg	5.0e11 vg
Hexb ^(-/-)	9	Dual: ICM high + IV medium	2.5e11 vg	7.5e11 vg	1.0e12 vg
Hexb ^(-/-)	7	Dual: ICM high + IV high	2.5e11 vg	2.5e12 vg	2.75e12 vg
Hexb ^(-/-)	9	IV only, low total dose	0	5.0e11 vg	5.0e11 vg
Hexb ^(-/-)	9	IV only, medium total dose	0	1.0e12 vg	1.0e12 vg
Hexb ^(-/-)	9	IV only, high total dose	0	2.75e12 vg	2.75e12 vg

Table 1. 10 cohorts of mice received concurrent infusions through both ICM and IV routes. There were 3 possible infusates for the ICM route (vehicle, low, or high vector dose) and 6 possible infusates (vehicle, 5 different vector doses) for the IV route.

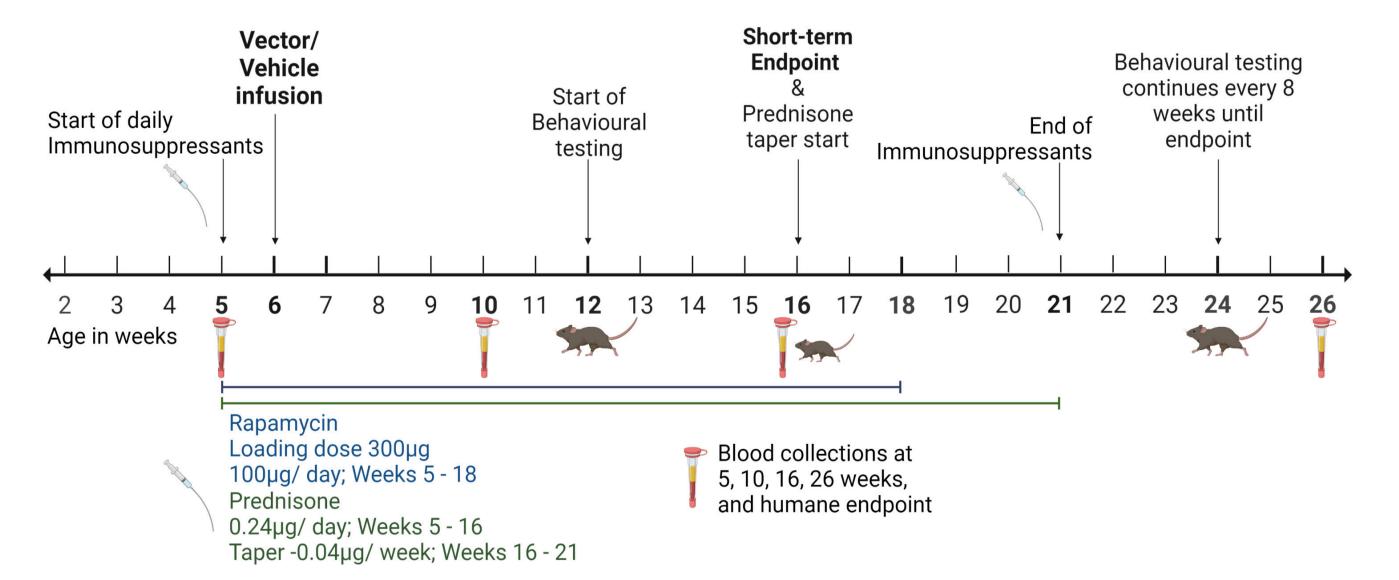


Figure 1. Timeline of Experimental Mice. Baseline blood collection and start of daily immunosuppressant regimen at 5-weeks. Administration of scAAV9-HexM or Vehicle infusions at 6-weeks. Immunosuppressant regimen maintained until 18-weeks (Rapamycin) and 21-weeks (Prednisone, tapered). Bimonthly behavioural testing, and blood collections at specific time points until mice reached their humane endpoint. At termination, blood, gross organs, brain, and spinal cord were collected for analysis of GM2 ganglioside accumulation, vector copy number, Hex enzyme activity, cellular and humoral immune response, and histology.

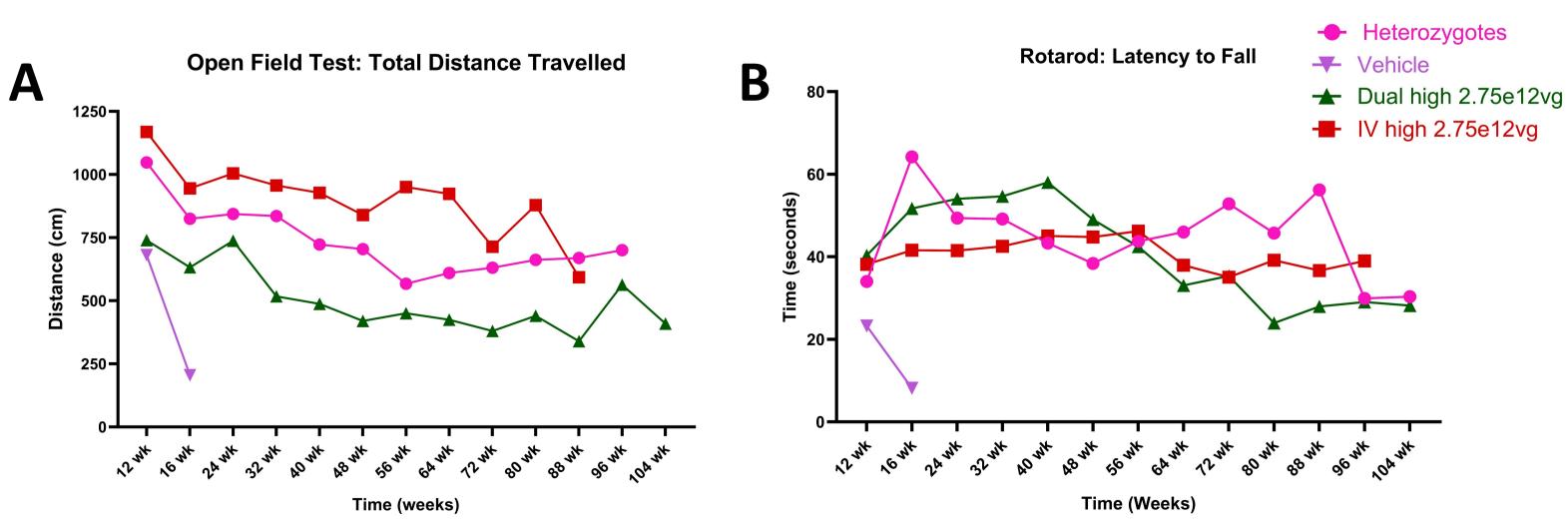


Figure 2. Behavioural Testing. Showed that the capabilities of IV high and Dual high treated mice were comparable to the heterozygous mice. (A) Open Field Testing showed no significant differences between cohorts for their Total Distance Travelled. Nor for the other parameters, not shown: Resting time, Mean speed, and Maximum speed. (B) Rotarod testing likewise showed no significant difference regarding their Latency to Fall, nor in the parameters not shown: Distance and End RPM.

NEW HOPE RESEARCH • FOUNDATION





Key Findings and Conclusions:

- The survival proportions show a strong dose-response and a clear benefit to splitting the dose between the ICM and IV routes when compared to the equivalent total dose in the IV-only cohorts (Figure 5).
- This study has shown up to a >6-fold increase in mean survival following scAAV9-HEXM vector infusion in six-week-old Sandhoff mice with survival of the longest-lived cohorts similar to the heterozygous control group (Figure 5).
- The longest-lived treated cohorts (Dual high and IV high) showed consistent behavioural performance over a two-year lifespan with no observable differences to that of the heterozygote control group (Figure 2).
- Results of the GM2 ganglioside accumulation assays echo the survival of the respective cohorts, with significant differences seen in multiple treated cohorts (Figure 6). The greatest reduction in GM2 gangliosides is seen in the Dual high cohort, which likewise was the longest surviving cohort, surpassing even the heterozygous control group.
- The equivalent survival and behavioural outcomes to the heterozygous group, coupled with the biochemical results, indicate that in the two high-dose cohorts (Dual high and IV high) Sandhoff disease has been corrected.
- This novel gene therapy for Sandhoff and Tay-Sachs Disease has tremendous implications for improved survival and quality of life in a clinical setting.

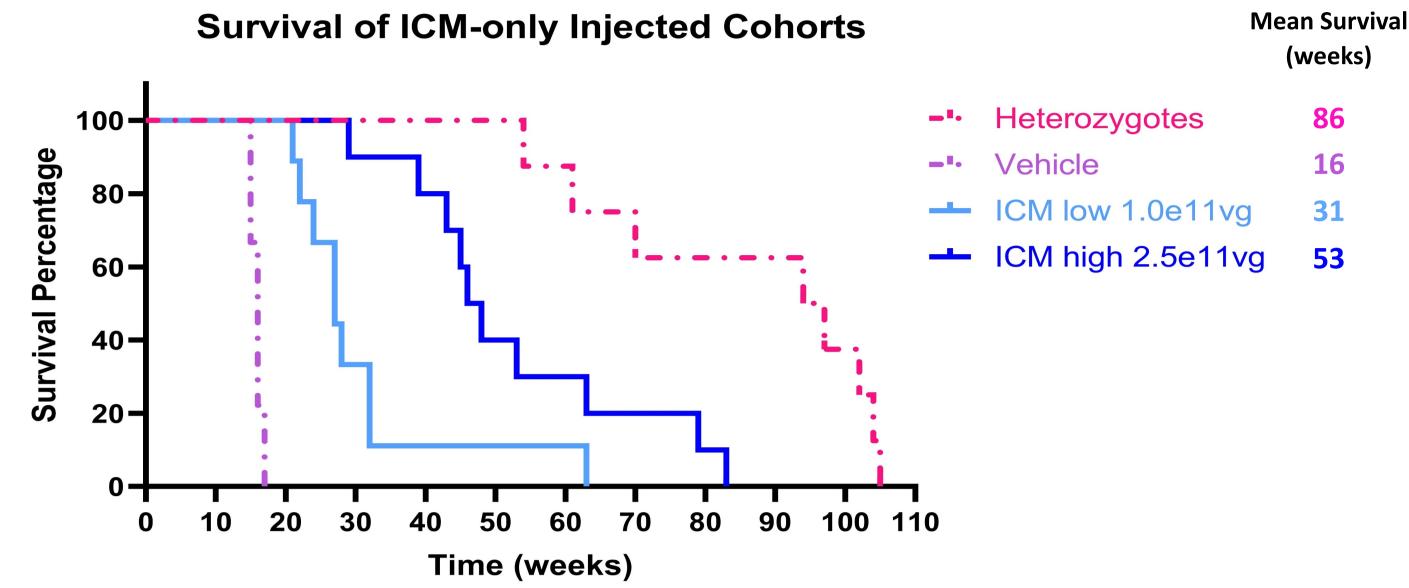


Figure 3. Survival of ICM-only Cohorts. Kaplan-Meier survival curves showing increased survival in both ICM-only cohorts. The ICM low (1.0e11vg) cohort shows a ~2-fold increase in mean survival, and the ICM high (2.5e11vg) cohort has a >3-fold increase compared to the vehicle-only control group.

Survival of IV-only Injected Cohorts

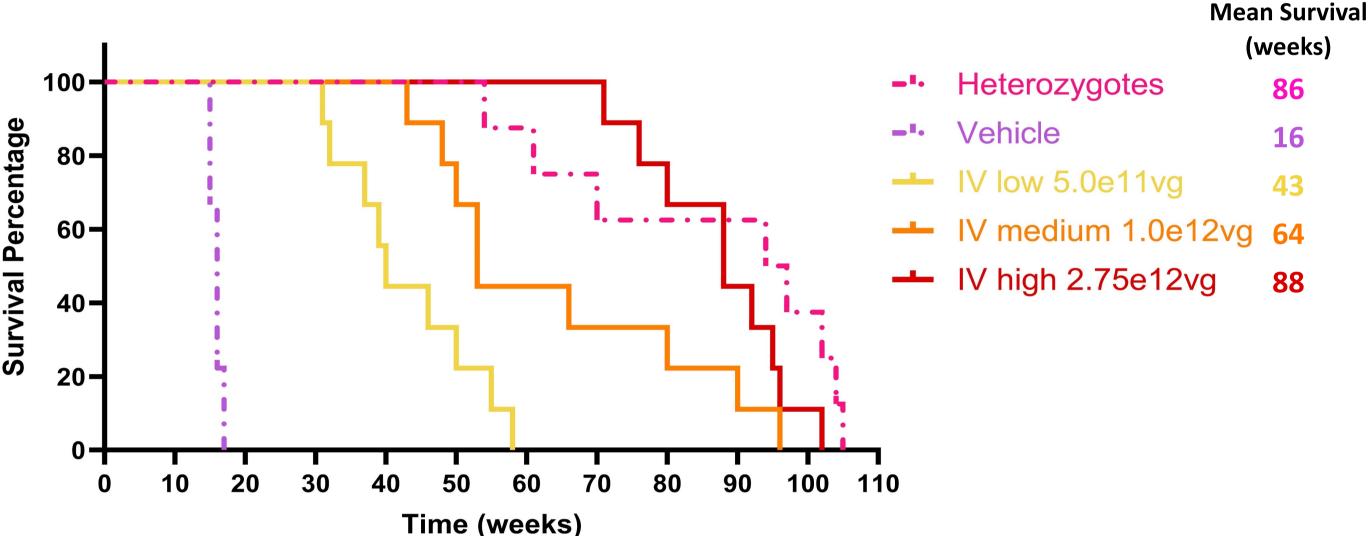


Figure 4. Survival of IV-only Cohorts. Kaplan-Meier survival curves showing a dose-response increase in survival in treatment groups compared to the vehicle-only control group (p < 0.001 in all cases). Mean survival increases almost 3fold in the IV low group (5.0e11vg), 4-fold in the IV medium group (1.0e12vg), and almost 6-fold in the IV high group (2.75e12vg). There is no significant difference between the IV high (2.75e12vg) cohort and the heterozygous cohort.

Survival of IV-only & Dual Injected Cohorts

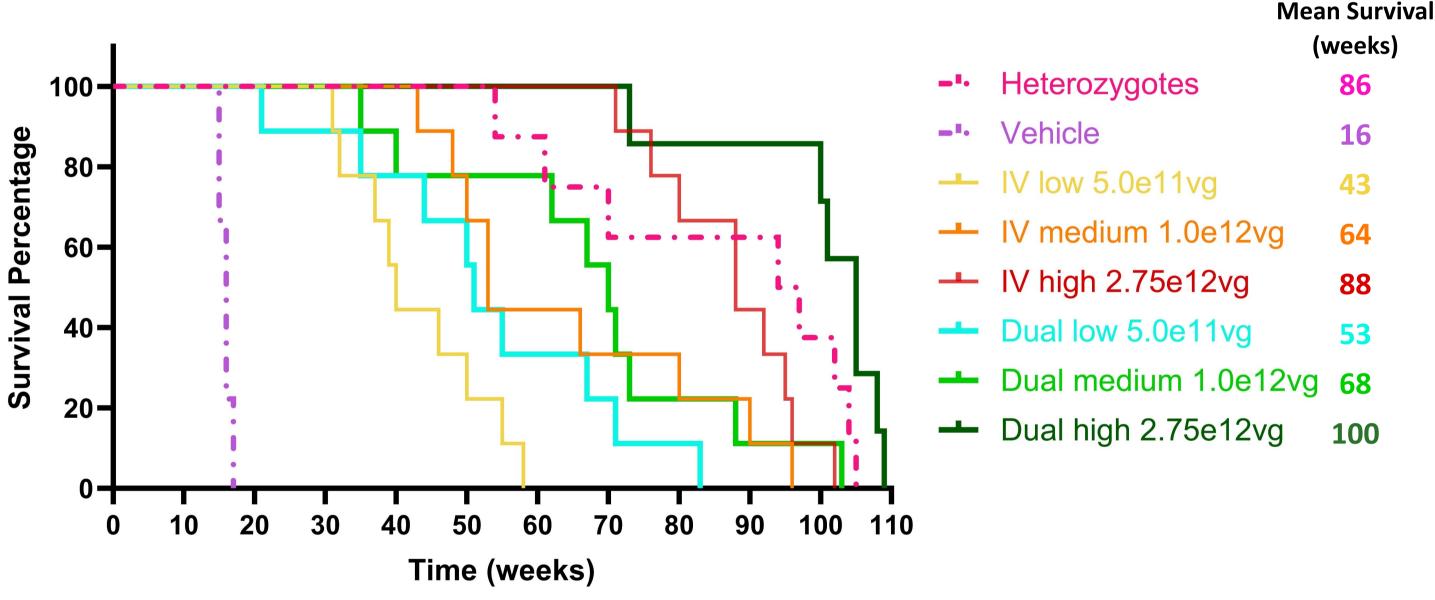


Figure 5. Survival of Dual-injected Cohorts Compared to IV-only. Kaplan-Meier survival curves showing a dose-response increase in survival in dual-treated groups compared to the vehicle-only control group (p < 0.001 in all cases). Mean survival increases >3-fold in the dual low (5.0e11vg) cohort, >4-fold in the dual medium cohort (1.0e12vg), and >6-fold in the dual high (2.75e12vg) cohort. Treatments split between the IV and ICM routes show a greater increase in mean survival compared to the equivalent dose delivered entirely IV.

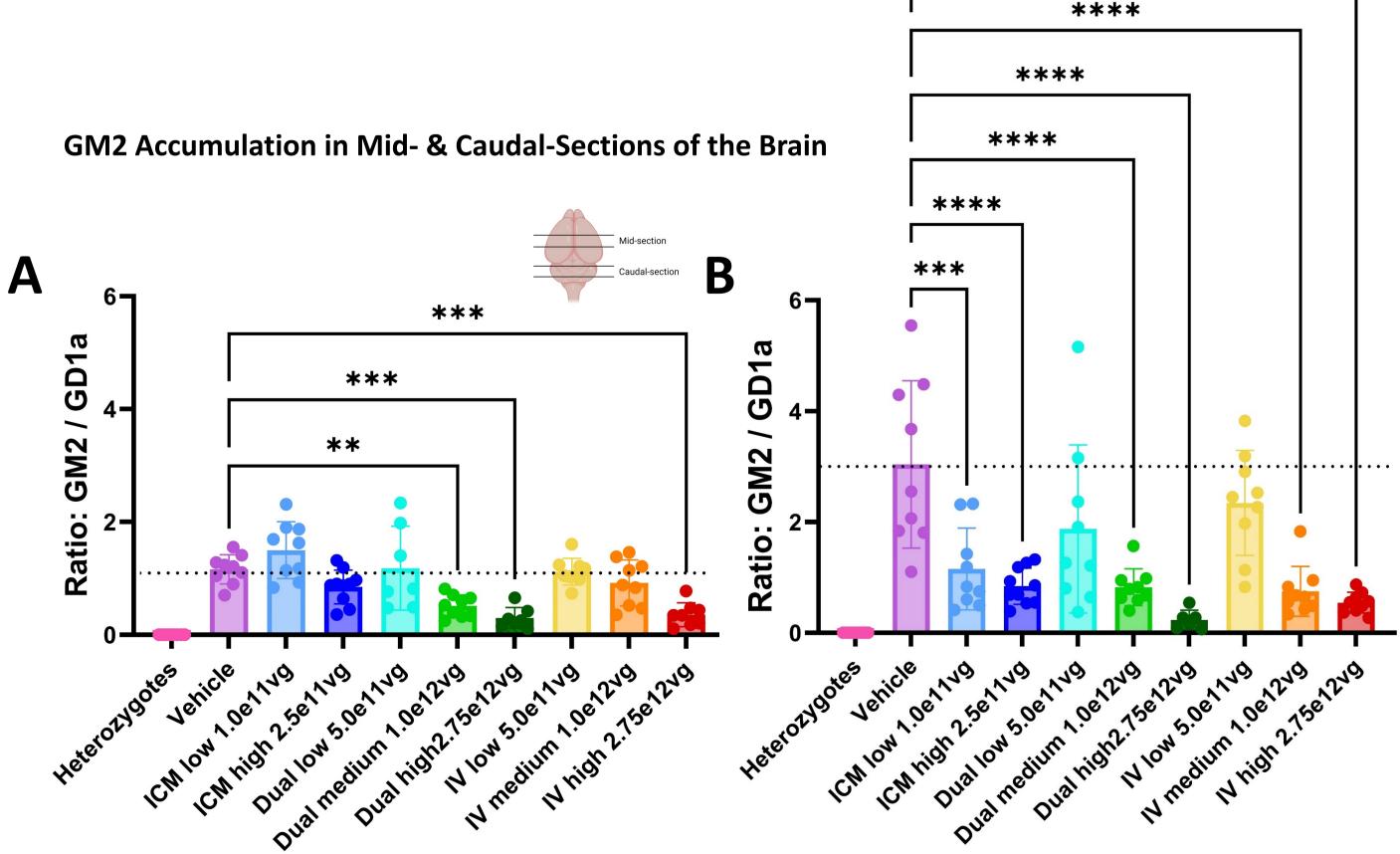


Figure 6. GM2 Ganglioside Accumulation. (A) In the mid-section of the brain, multiple treated cohorts show a significant decrease in accumulated GM2 gangliosides compared to vehicle-only controls, the greatest being Dual high with a >3.8-fold decrease (*** p < 0.001). (B) In the caudal-section of the brain, almost all treated cohorts have a significant decrease in GM2 accumulation compared to vehicle-only controls, the greatest being a >12.9-fold decrease in the Dual high cohort (**** p < 0.0001). Samples were collected at humane endpoint, at which stage GM2 ganglioside accumulation is expected to be at its highest.